

# Antibacterial Drug Discovery Using Phage-Validated, Novel Targets

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**1 Abstract**

**Background:** Viruses, and their interactions with their hosts, have played a crucial role in assessing many fundamental rules of life. Over billions of years of evolution, the viruses of subunits, called "bacteriophages" or "phages", have developed unique proteins enabling them to arrest or inhibit critical bacterial cellular processes to augment phage reproduction. To address concerns of the sinking global increase in antibiotic resistance, we have exploited the mechanisms of bacterial growth inhibition employed by these phages as tools to discover novel cellular targets for antibiotic discovery and development. Methods Using high-throughput bacteriophage genomics and functional genomics, we sequenced the genomes of 47 phages that infect three pathogens (*Staphylococcus aureus*, *Streptococcus pneumoniae* and *Pseudomonas aeruginosa*). The open reading frames (ORFs) of these phages were identified and screened for those that inhibited bacterial growth when induced in their hosts. These phage-derived inhibitory ORFs were used as tools to systematically identify their interacting partners in bacteria. Results A variety of targets which were biologically validated by phages and essential for bacterial growth were discovered by our technology platform. High-throughput screens (HTS) campaigns were carried out against some of these targets which are involved in DNA replication and RNA transcription. Small molecule leads were identified. These compounds inhibited bacterial growth and may mimic the effect of the phage ORFs on their cognate bacterial targets. Lead optimization is under way. Conclusion By taking advantage of what we have learnt from nature, our technology platform plays an important role in the battle against increasing antibiotic resistance.

**2 Introduction**

With its unique proprietary phage genomics platform, PhageTech Inc. identifies and exploits the natural mechanisms developed by bacterial viruses (phages) that lead to the death of their bacterial hosts, including human pathogens. PhageTech has assembled a large collection of phages and a unique proprietary database of the genomes of phages that kill *Staphylococcus aureus*, *Streptococcus pneumoniae* and *Pseudomonas aeruginosa*. Functional genomics tools are used to identify candidate growth inhibitory proteins (ORFs) from the phages; proteomics tools are then applied to identify the cognate bacterial targets of these phage-derived growth inhibitory proteins. Growth inhibitory protein families and their bacterial targets have been identified. HTS against these targets has identified a number of novel small molecule compounds that may mimic the phage-derived inhibitory proteins in their antibacterial effects. Different from phage therapy, this approach supports the development of new classes of small molecule antibiotics against phage-selected bacterial targets involved in essential bacterial functions.

**3 Major Approaches to Antibacterial Discovery**

- Screening of candidate molecules in whole-cell based assays for antibacterial activity
- Analoging of known drugs or drug candidates with the goal of creating newer or improved versions
- Discovering of novel compounds for known therapeutic targets with established mode of action
- Target-based drug discovery via bacterial targets selected by phages - PhageTech approach

Key advantages of the PhageTech drug discovery platform include:

- speed of analysis of small phage genomes
- access to diversified targets
- inherent validation of the bacterial targets by phages

**4 Summary of PhageTech's Genomics and Functional Genomics Programs**

Bacterial pathogen	Phages collected	Genomes sequenced	ORFs screened	Inhibitor families
<i>S. aureus</i>	150	27	984	31
<i>S. pneumoniae</i>	50	8	264	5
<i>P. aeruginosa</i>	70	11	500	13
<b>Total</b>	<b>270</b>	<b>46</b>	<b>1728</b>	<b>49</b>

**5 ORF Affinity Approach to Target ID**

Affinity chromatography of bacterial lysate over immobilized inhibitory ORF

↓

Tryptic peptide mapping, mass spectrometry of eluted proteins

↓

Target identification

↓

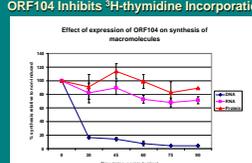
Target validation

- Confirm ORF-Target Interaction
- Confirm target essentiality
- Determine target function

**6 Identification of R1 involved in DNA Synthesis as a *S. aureus* Target of ORF104**



**7 ORF104 Inhibits <sup>3</sup>H-thymidine Incorporation**



**8 Validation of Bacterial Targets from the Proteomics Platform**

- Several targets were identified from *S. aureus* lysates using immobilized inhibitory phage ORFs
- Direct interactions between inhibitory phage ORFs and their cognate bacterial targets were confirmed by:
  - Mass two-hybrid analysis
  - Far western (protein blotting)
  - ELISA
  - Time-resolved FRET (TR-FRET)
  - Fluorescence polarization (FP)
- These bacterial targets are:
  - essential
  - attractive targets for antibiotic discovery

**9 *S. aureus* DNA and RNA Synthesis Proteins Identified by Phage Inhibitory ORFs**

Representative of inhibitory ORF family	ORF size (aa)	Bacterial target identified	Essentiality of target
ORF104	52	R1 (DNA)	Essential
ORF105	287	R1 (DNA)	Essential
ORF225	59	R2 (DNA)	Essential
ORF169	74	R2 (DNA)	Essential
ORF240	58	R2 (DNA)	Essential
ORF078	71	R9 (DNA)	Essential
ORF27	198	R12 (RNA)	Essential

**10 Summary of Assay Technology**

Assay	Players	Type	Principle	Readout
FP	Target / ORF	Protein-protein	Target polarizes ORF receptor	Fluorescence
TR-FRET	Target / ORF	Protein-protein	FRET with proximity of donor to acceptor	Fluorescence
TCA	Target	Enzyme	Label incorporation into product	Radioactivity

**11 Screen Strategy**

Small molecule compounds

↓ Primary screening

Actives

↓ Confirmatory screening

Confirmed actives

↓ Dose response testing

Hits

↓ In vitro functional assay, if available

Confirmed hits

↓ Susceptibility testing and compound profiling

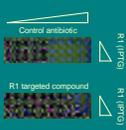
Lead

**12 Hit Compounds Identified Against *S. aureus* Targets**

Target	Assay	Library	Actives	Confirmed actives	Hits (IC <sub>50</sub> = 10µM or less)	Confirmed Hits (MIC = 16µg/ml or less)
R1 (DNA)	FP	125 K	1311	156	73	16
R9 (DNA)	TR-FRET	125 K	932	464	90	24
R12 (RNA)	TCA	125 K	830	193	73	36

**13 Mechanism of Action Studies Using R1 Titrating System**

- Use *S. aureus* strain with R1 expression under inducible promoter
- Susceptibility testing for ORF (R1)-derived small molecule compounds
- Assay in the presence of increasing concentrations of inducer IPTG
- Antibiotics against known targets other than R1 as controls



**14 Properties of ORF104/R1-Derived Small Molecules**

Small molecule compound	ORF104-R1 interaction % (µM)	MIC against <i>S. aureus</i> (µg/mL)	DNA synthesis IC <sub>50</sub> (µg/mL)	RNA synthesis IC <sub>50</sub> (µg/mL)	Fold effect in MIC under R1 regulated expression (100/25 µM IPTG)
1	1.5	0.125	0.8	1.5	4
2	0.6	1	2.4	6.5	8
3	4.8	16	1.5	13.7	8

- Data suggest these compounds kill bacteria by targeting R1
- Confirmed hits are now the focus of SAR / chemistry programs

**15 Activities of One Lead Series Against R12**

Compound	TCA IC <sub>50</sub> (µM)	MIC (µg/ml)			
		<i>S. aureus</i>	<i>S. pneumoniae</i>	<i>E. coli</i>	<i>E. coli</i> hsdR
PT11010480	7	0.5	4	>32	0.5
PT99000019	9	4	4	>32	1
PT99000021	2	1	4	>32	1
PT99000029	4	4	4	>32	1
PT99000030	1	1	4	32	0.25
PT99000031	2	2	1	32	0.5

- These compounds kill both Gram positive and Gram negative bacteria

**16 Efflux Pumps Lead Compounds Vs Known Antibiotics**

Compound	MIC (µg/ml)		
	<i>S. aureus</i>	<i>S. aureus</i> efflux	<i>S. aureus</i> efflux
PT11010480	0.5	2	0.5
PT99000030	1	0.5	0.031
PT99000031	2	2	0.062
Norfloxacin	1	32	4
TPP	8	32	32
Erythromycin	0.25	0.25	0.25

- These compounds are not substrates of major efflux pumps

**17 MRSA Lead Compounds Vs Known Antibiotics**

Compound	MIC (µg/ml)				
	<i>S. aureus</i>	MRSA-1	MRSA-2	MRSA-3	MRSA-4
PT11010480	0.5	0.25	1	1	8
PT99000030	1	0.125	0.5	0.5	4
PT99000031	2	0.5	1	2	16
Oxacillin	0.25	>32	>32	>32	>32
Vancomycin	1	1	1	2	1
Erythromycin	0.25	>64	>64	>64	>64
Norfloxacin	1	0.5	0.5	32	0.5

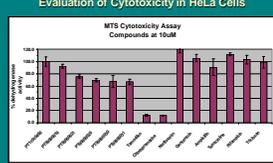
- These compounds are active against MRSA

**18 Effect of Serum Proteins on Activity of Compounds**

Compound	MIC for <i>S. aureus</i> (µg/ml)		
	CAMBIB	+25% mouse serum	+25% human serum
PT11010480	0.25	0.5	0.5
PT99000019	0.25	0.5	0.5
PT99000021	0.125	0.25	0.25
PT99000029	0.125	0.25	0.25
PT99000030	<0.125	<0.125	<0.125
PT99000031	<0.125	0.125	0.25

- These compounds are active even in the presence of serum

**19 Evaluation of Cytotoxicity in HeLa Cells**



- Cytotoxicity was also evaluated in primary hepatocytes

**20 Summary**

- PhageTech has sequenced the genomes of 47 phages of *S. aureus*, *S. pneumoniae* and *P. aeruginosa*
- 49 families of phage-derived antimicrobial ORFs were identified
- Several biologically validated, essential bacterial targets, including those required for DNA and RNA synthesis in *S. aureus*, have been discovered
- Proprietary ORF-target binding assays and enzymatic assays have been developed around these targets
- Target titration systems have been developed and will be used for demonstration of mechanism of action
- Diverse libraries of small molecules are being screened for inhibitors and selected compounds are being optimized
- Some lead compounds show promising antibacterial profiles against Gram positive and Gram negative bacteria including drug resistant organisms
- Efficacy of lead compounds will be evaluated in animal models