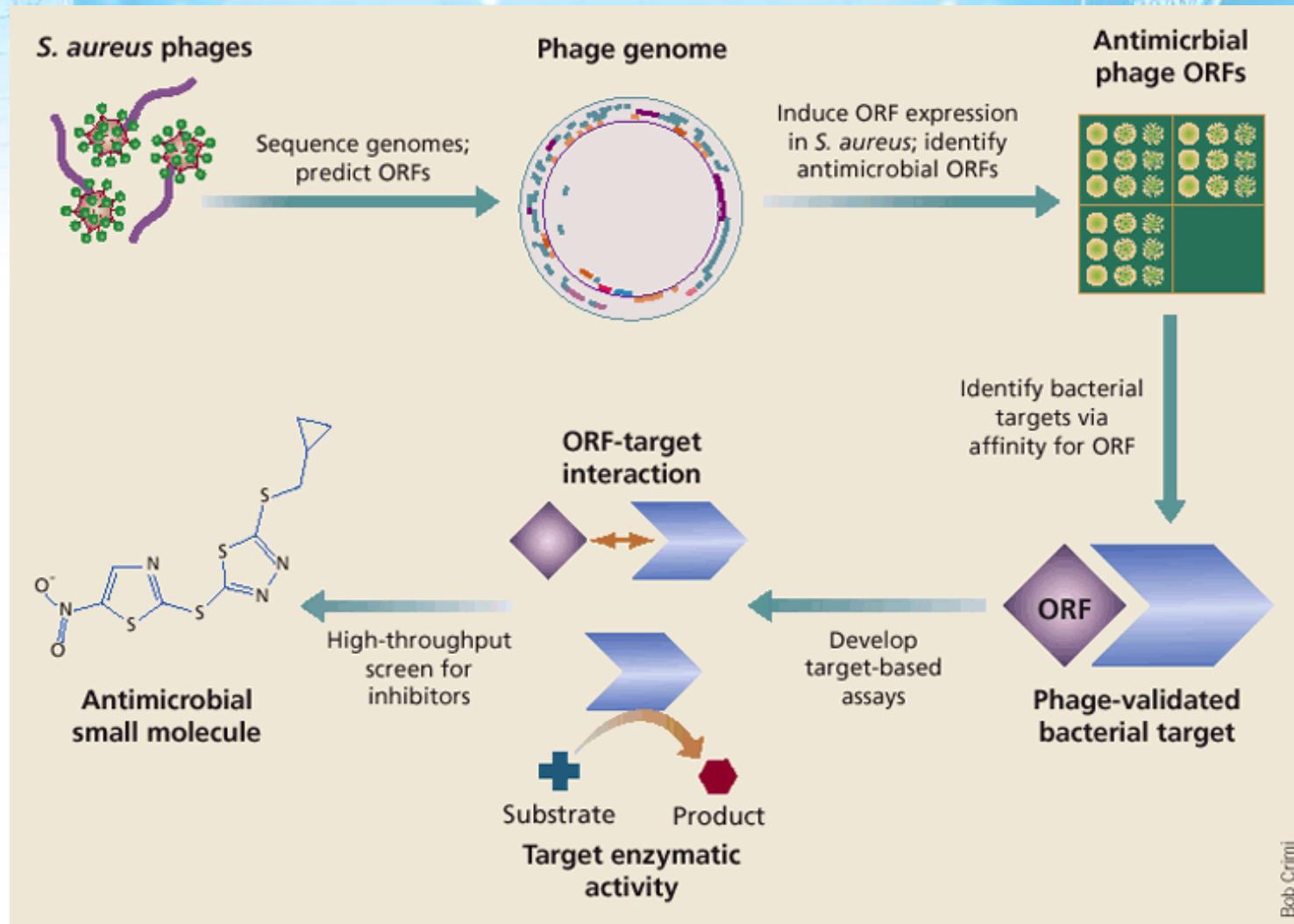


Phage-Inspired Discovery of Small-Molecule Antibiotics

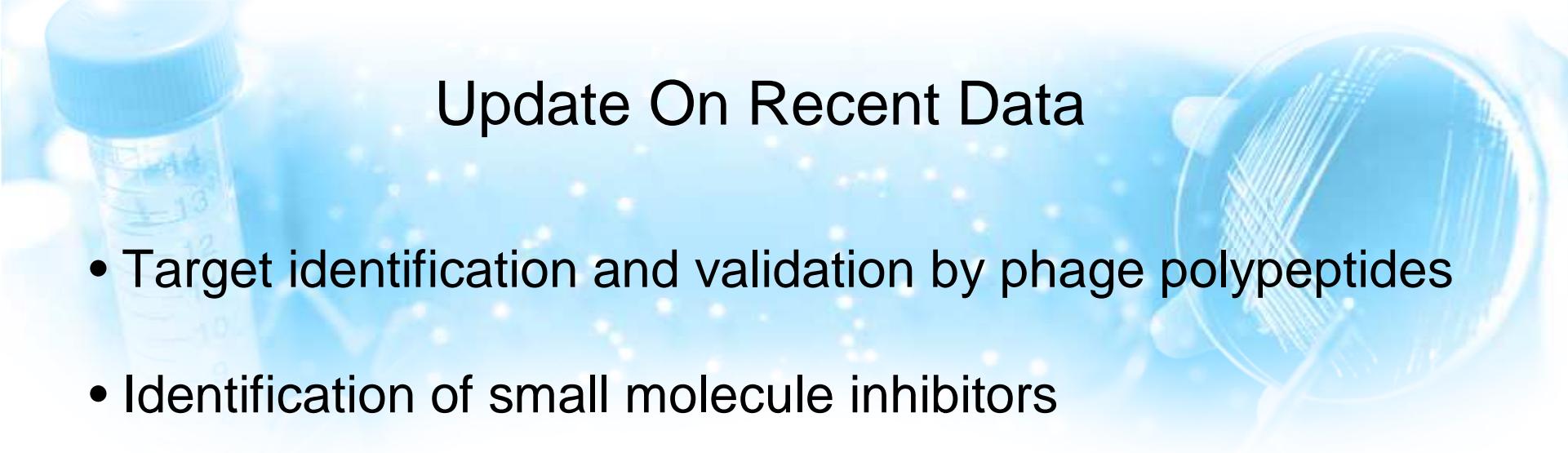
May 18, 2006

Phage-Inspired Drug Discovery Approach



Liu et al., 2004. *Nature Biotech.* 22:185

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Update On Recent Data

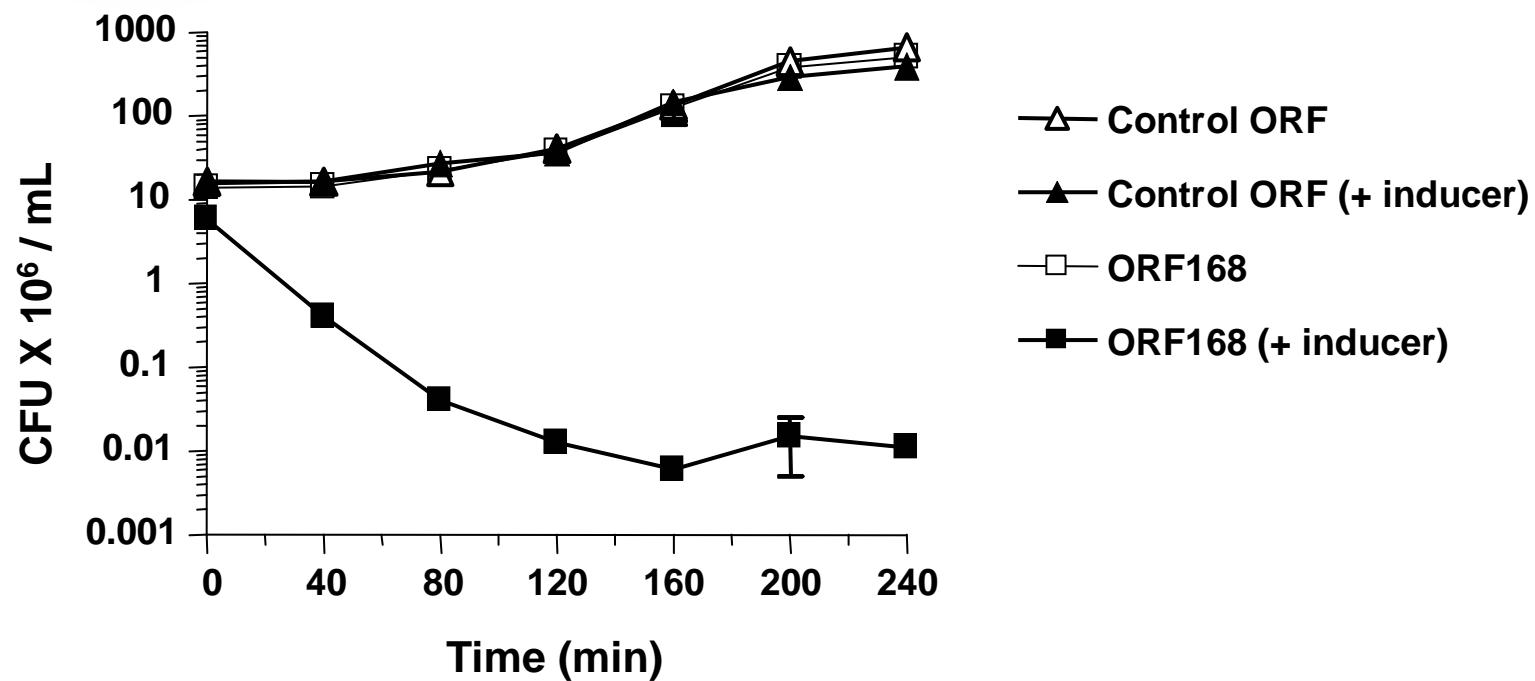
- Target identification and validation by phage polypeptides
- Identification of small molecule inhibitors
- Optimization of inhibitors via medicinal chemistry
- Two examples:
 - *S. aureus* DNA replication target: DNA polymerase β subunit
 - *S. aureus* transcription target: primary sigma factor RpoD



S. aureus DNA replication target: DNA polymerase β subunit

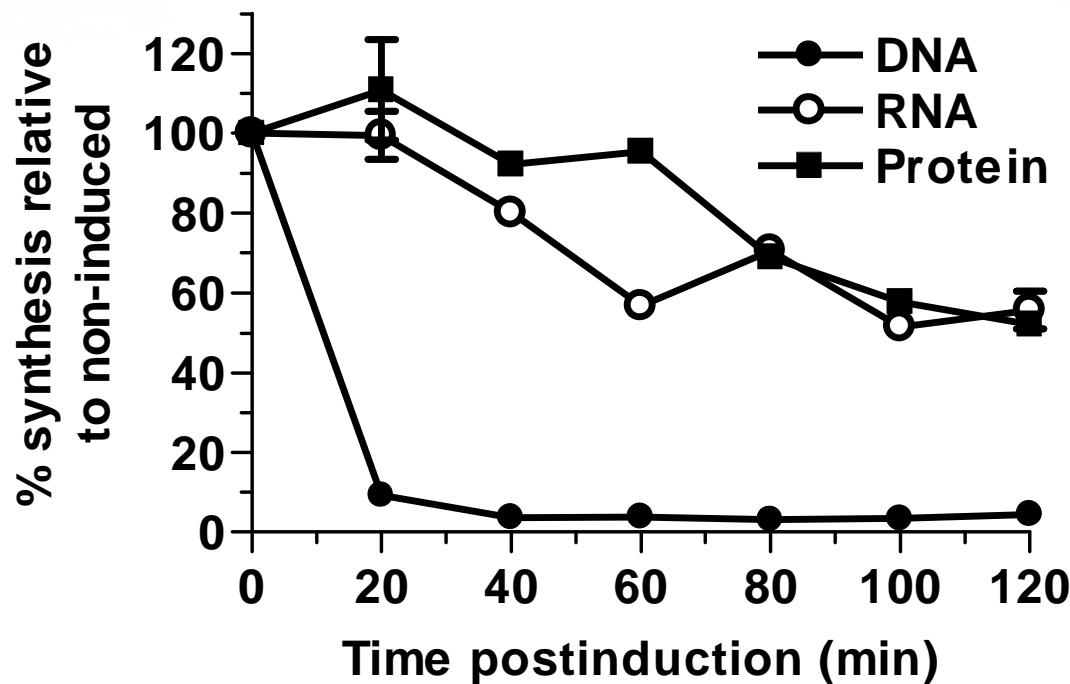
Expression of ORF168 in *S. aureus* is Bactericidal

- Dot screening → phage open reading frames that inhibit growth when expressed within *S. aureus*
- Broth assay → rapid killing kinetics of selected phage ORFs:

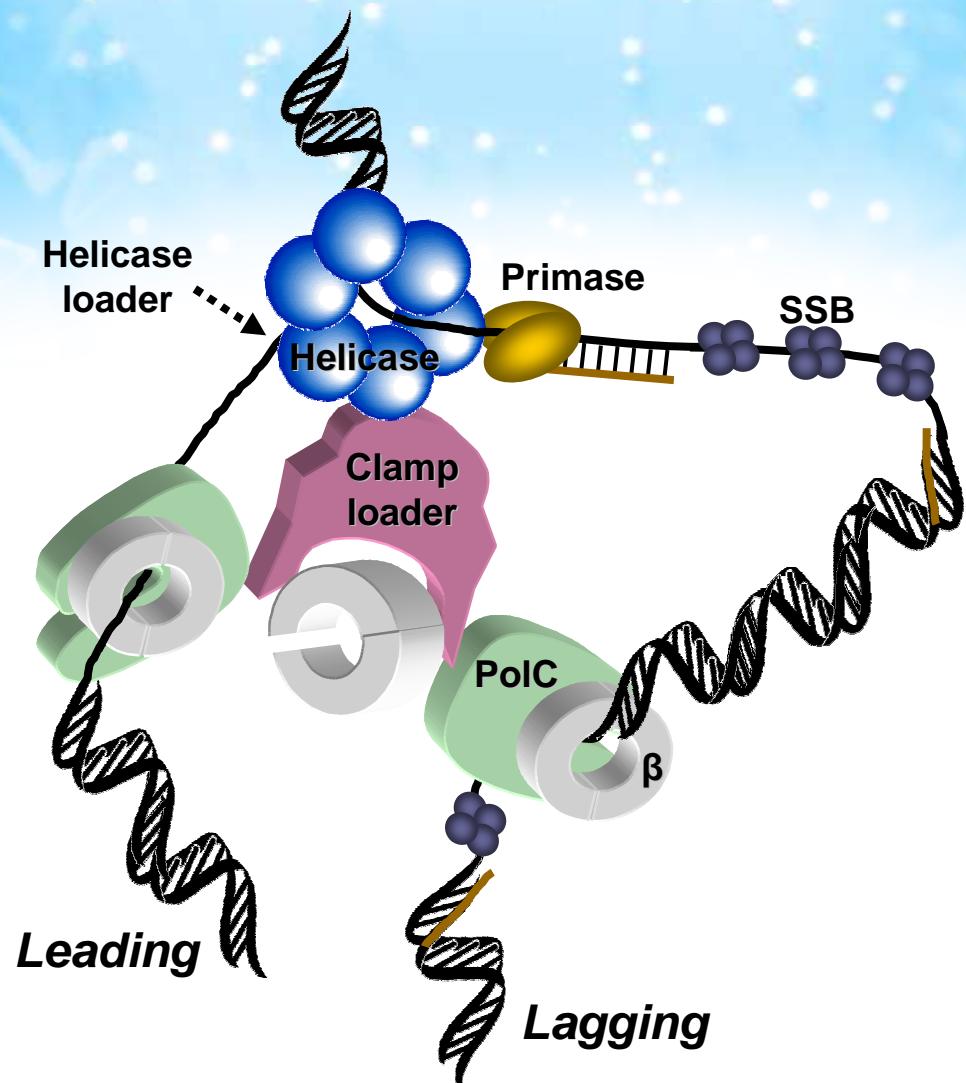


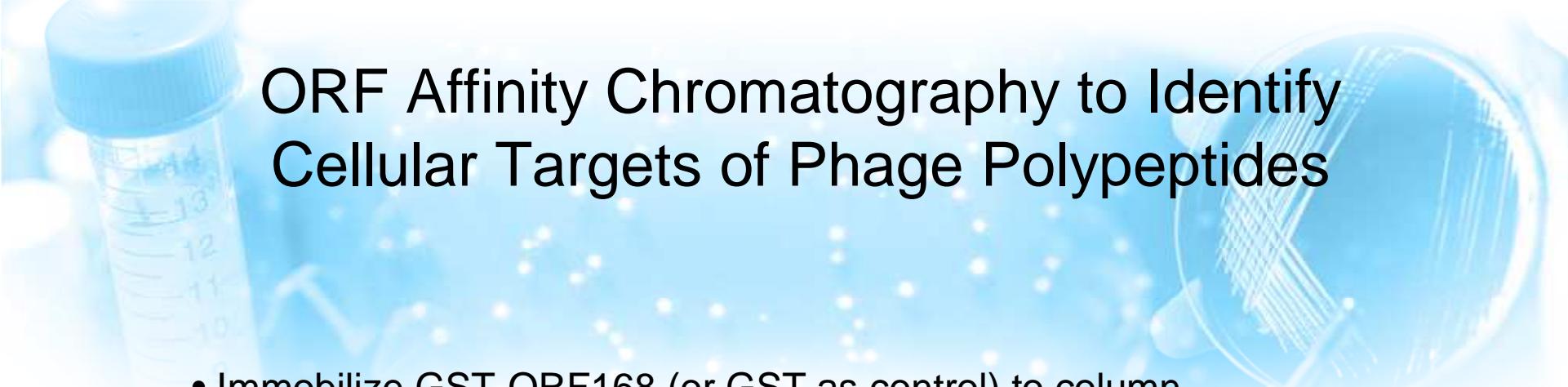
Expression of ORF168 in *S. aureus* Selectively Inhibits DNA Synthesis

- Macromolecular synthesis assay in *S. aureus* → selectivity of inhibition:



The Bacterial DNA Replication Machinery Offers Essential, Under-Exploited Targets

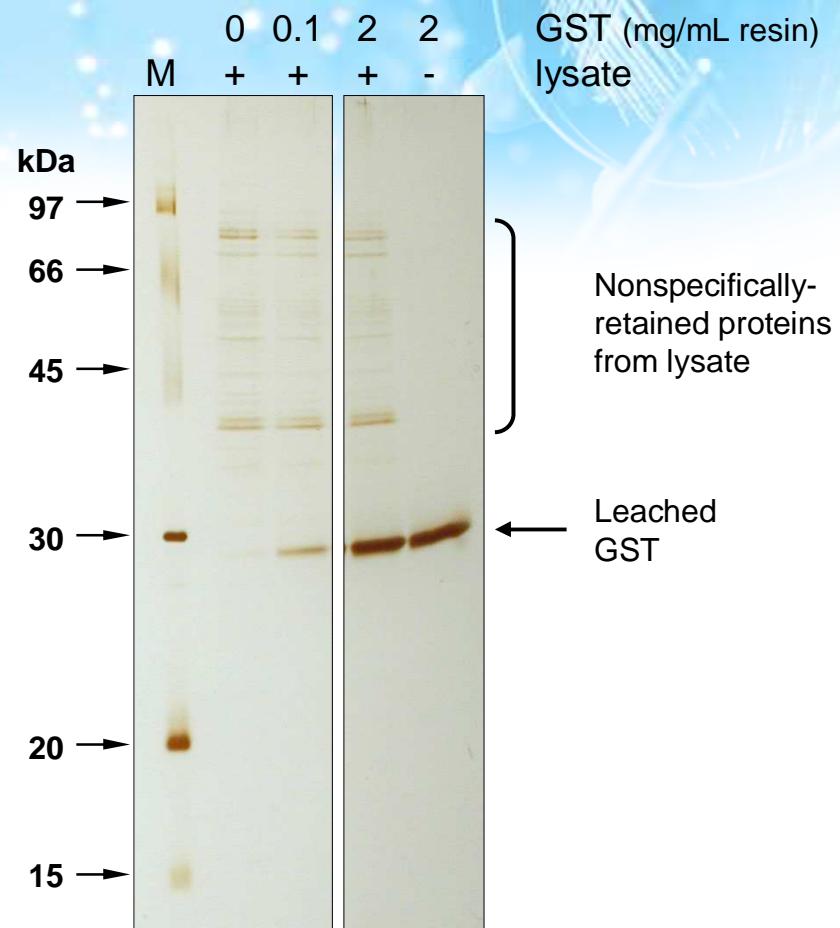




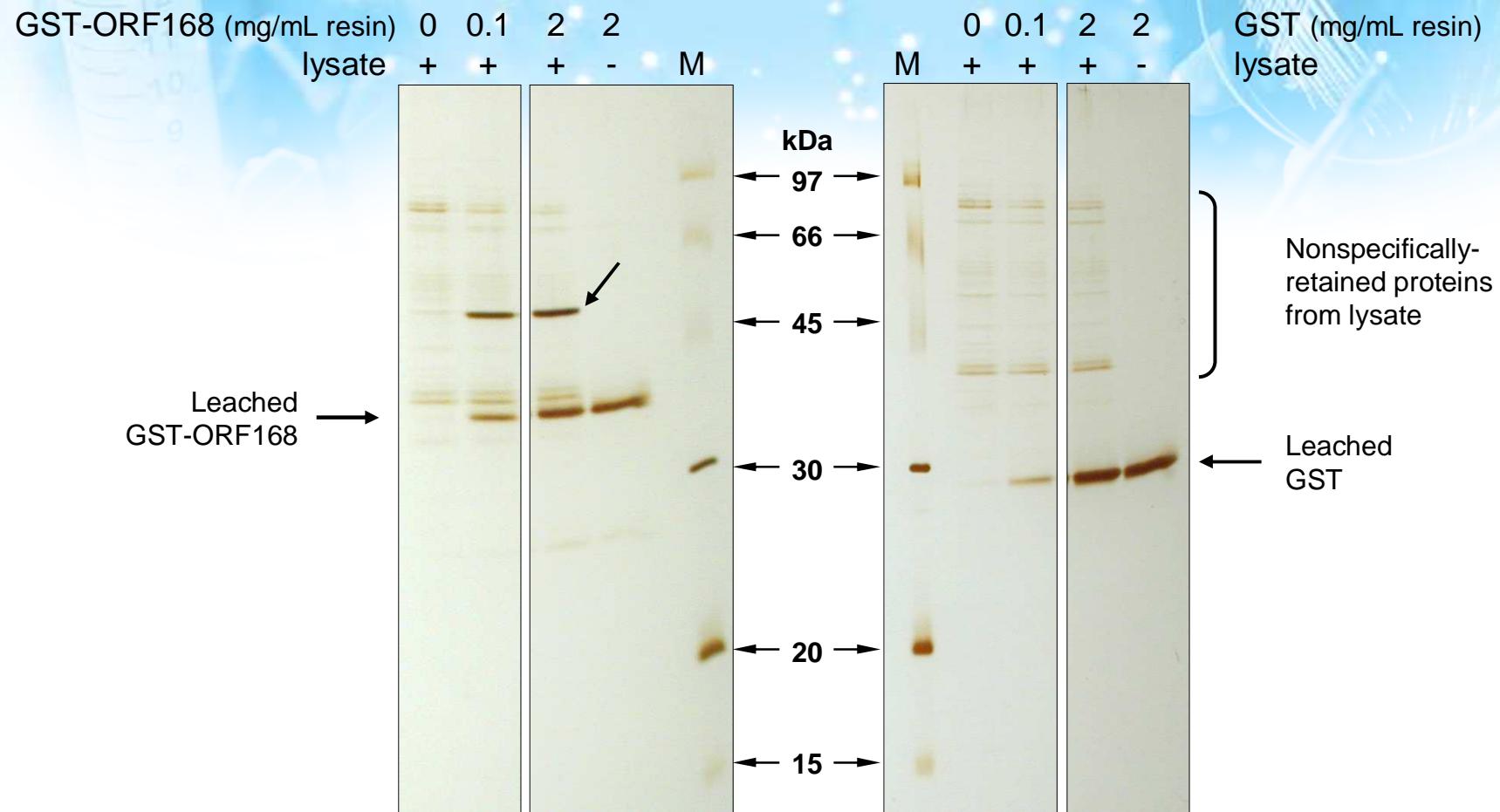
ORF Affinity Chromatography to Identify Cellular Targets of Phage Polypeptides

- Immobilize GST-ORF168 (or GST as control) to column
- Apply *S. aureus* extract (or buffer as control)
- Wash, elute retained proteins with 1% SDS
- SDS-PAGE, silver staining
- Excise bands of interest and digest with trypsin
- Determine masses of tryptic peptides by MALDI-ToF mass spectrometry

ORF Affinity Chromatography to Identify Cellular Targets of Phage Polypeptides

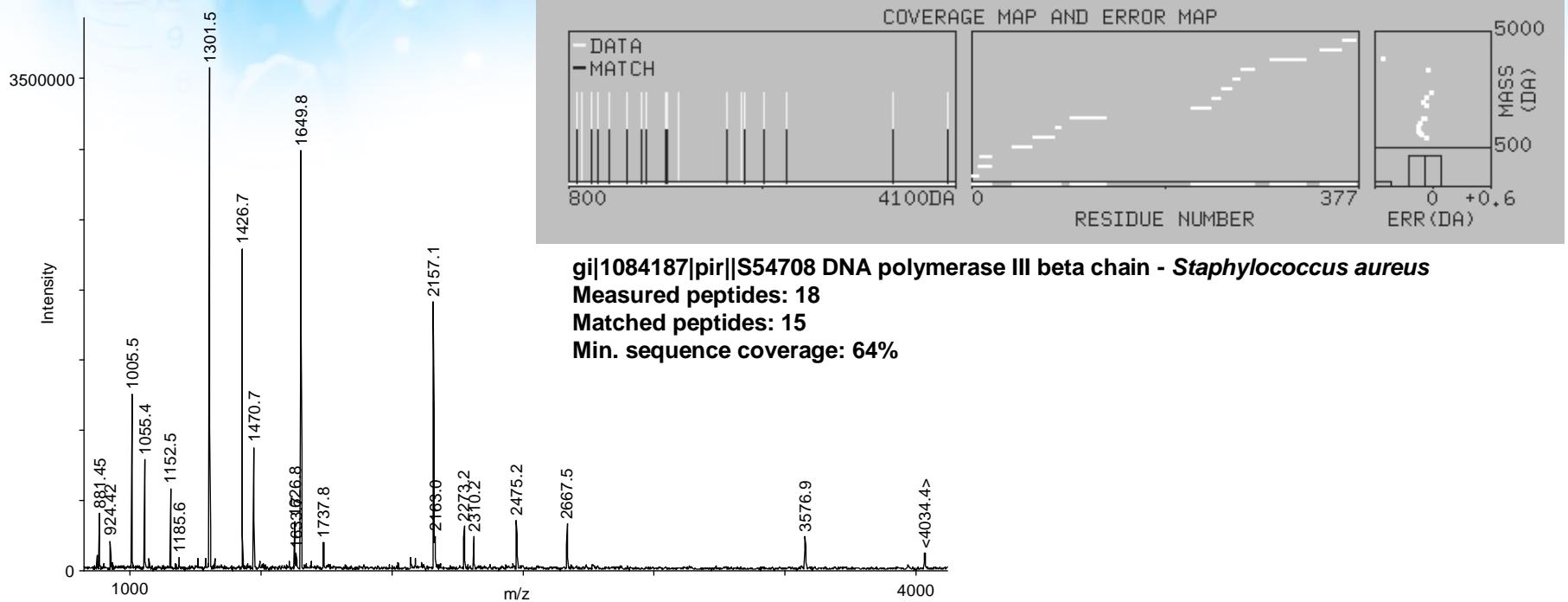


ORF168 Affinity Chromatography Identifies a ~45 kDa Prey Protein from *S. aureus* lysate



- One candidate interacting protein was eluted specifically from the ORF168 affinity column

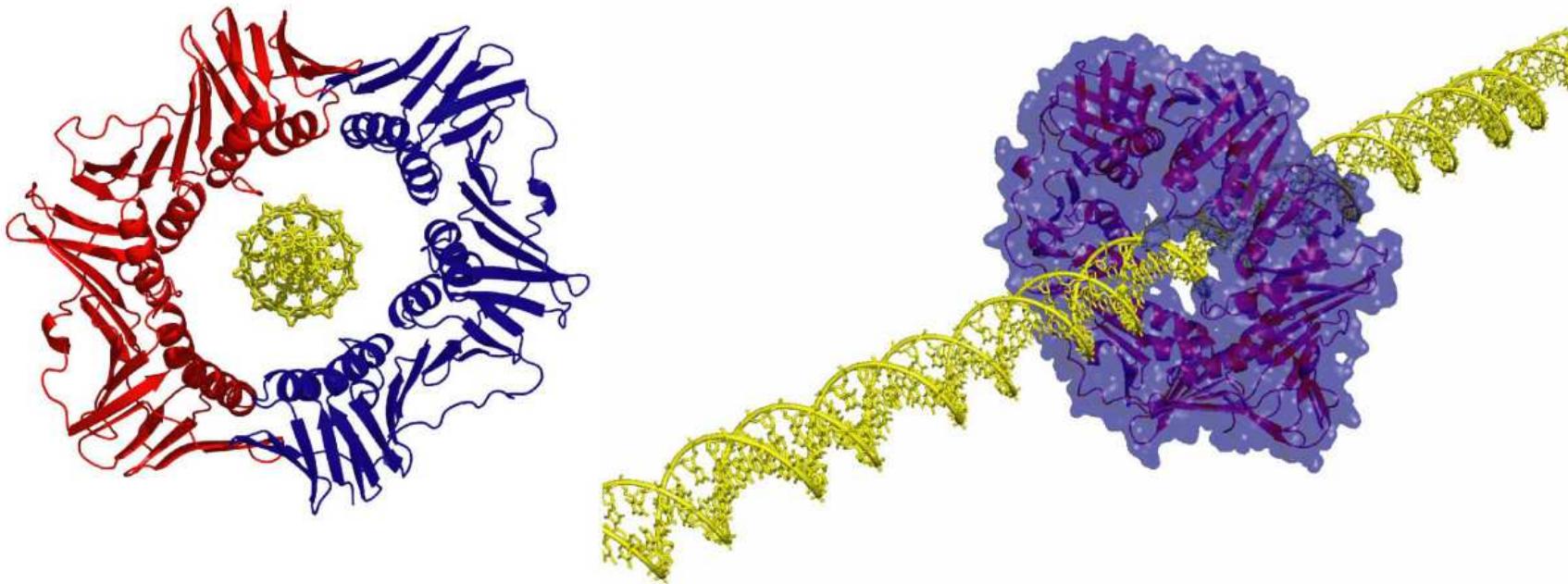
Tryptic Fingerprint Identifies *S. aureus* DNA Pol β as the Target of ORF168



- Tryptic peptide mass spectrum identifies the target of ORF168 as the β subunit of *S. aureus* DNA Polymerase C

Structure of DNA Pol β from *Streptococcus pyogenes*

- Three-domain structure in protomer
- Homodimer shares overall topology with other bacterial DNA sliding clamps despite sequence divergence (e.g. 23% a.a. identity with *E. coli* β)



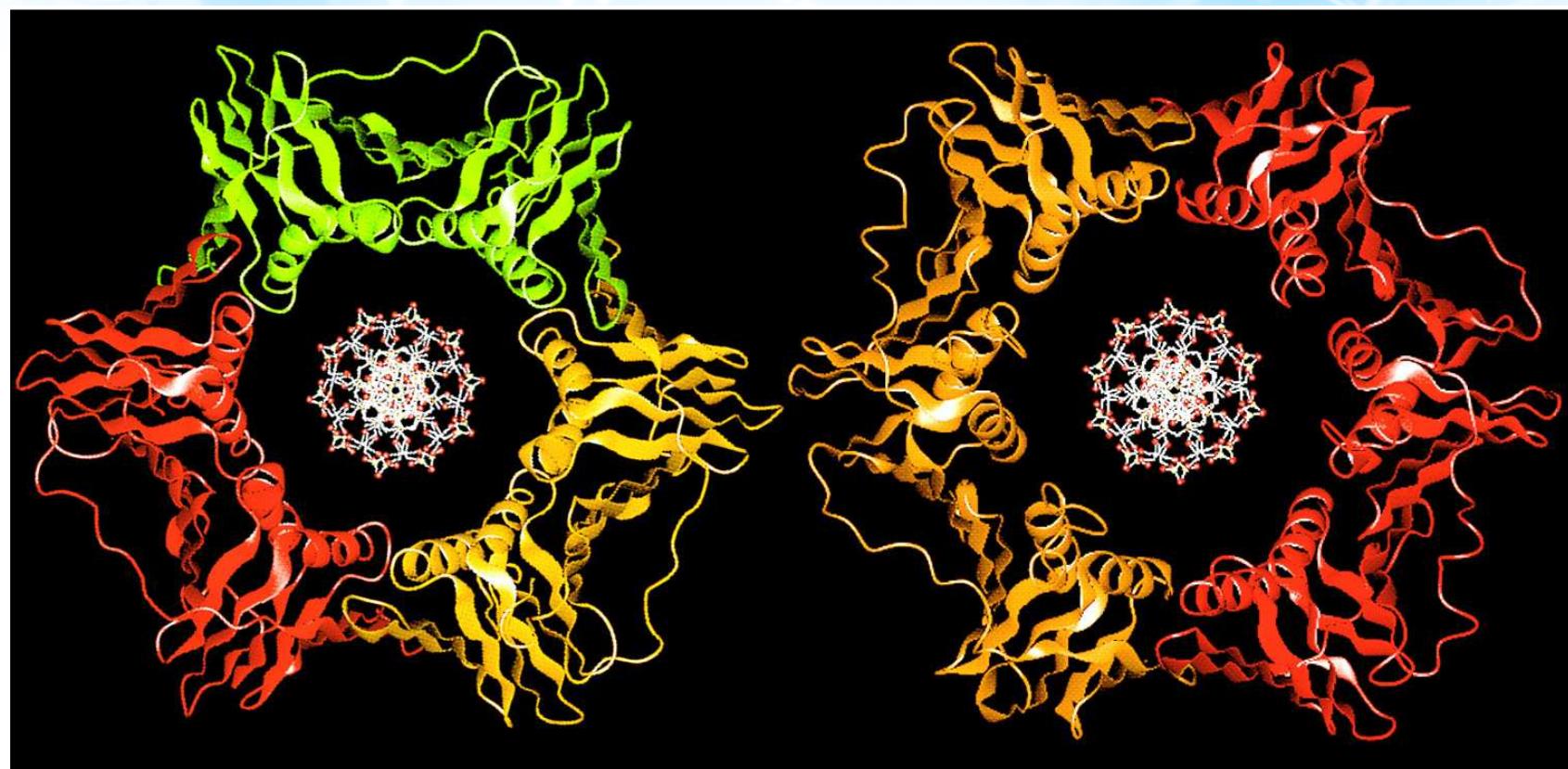
Argiriadi et al., 2006. BMC Struct. Biol. 6:2

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DNA Sliding Clamps Share Mechanistic Similarity Despite Divergence in Sequence

Yeast PCNA

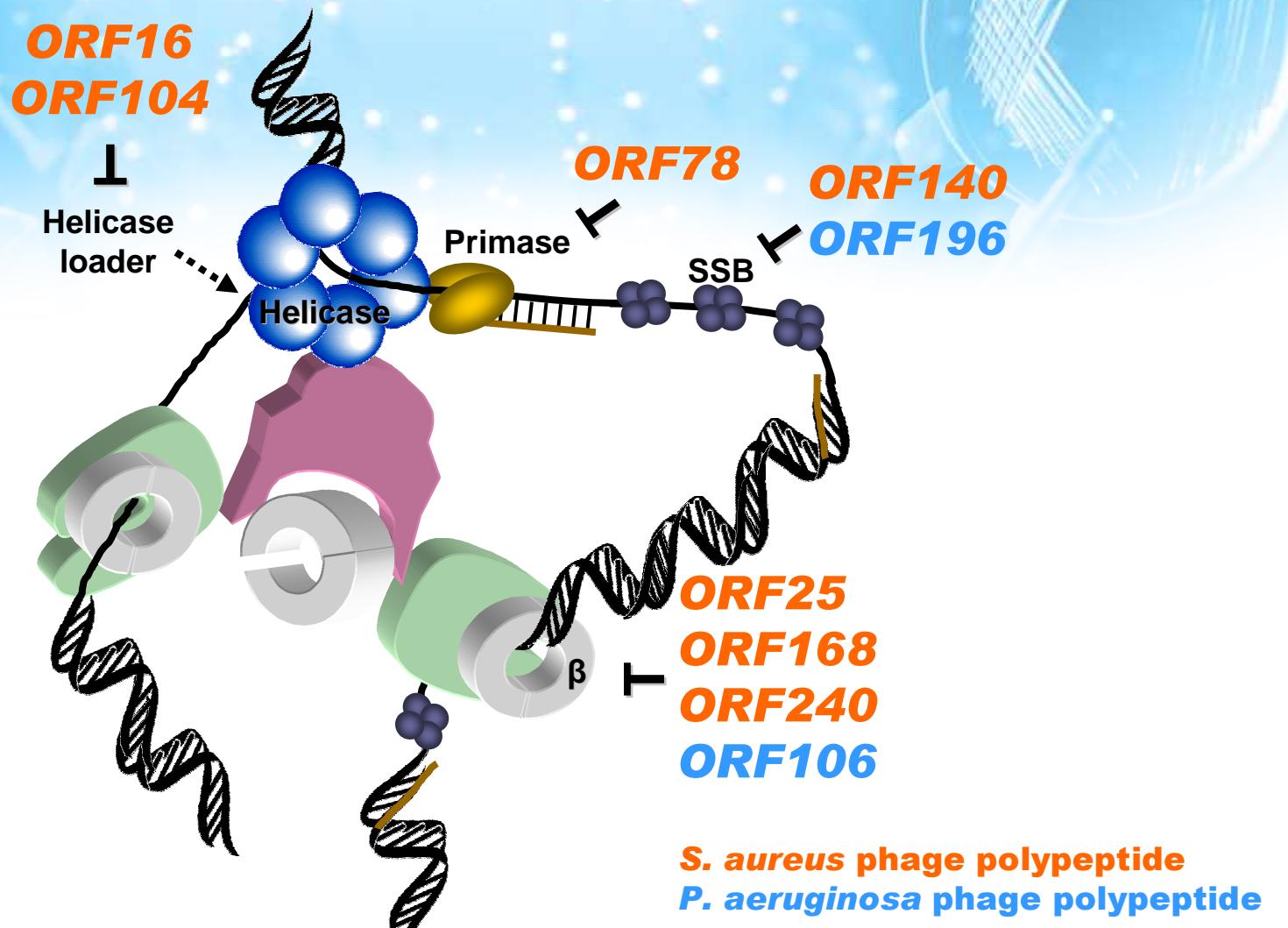
E. coli β



Krishna et al., 1994. Cell 79:1233

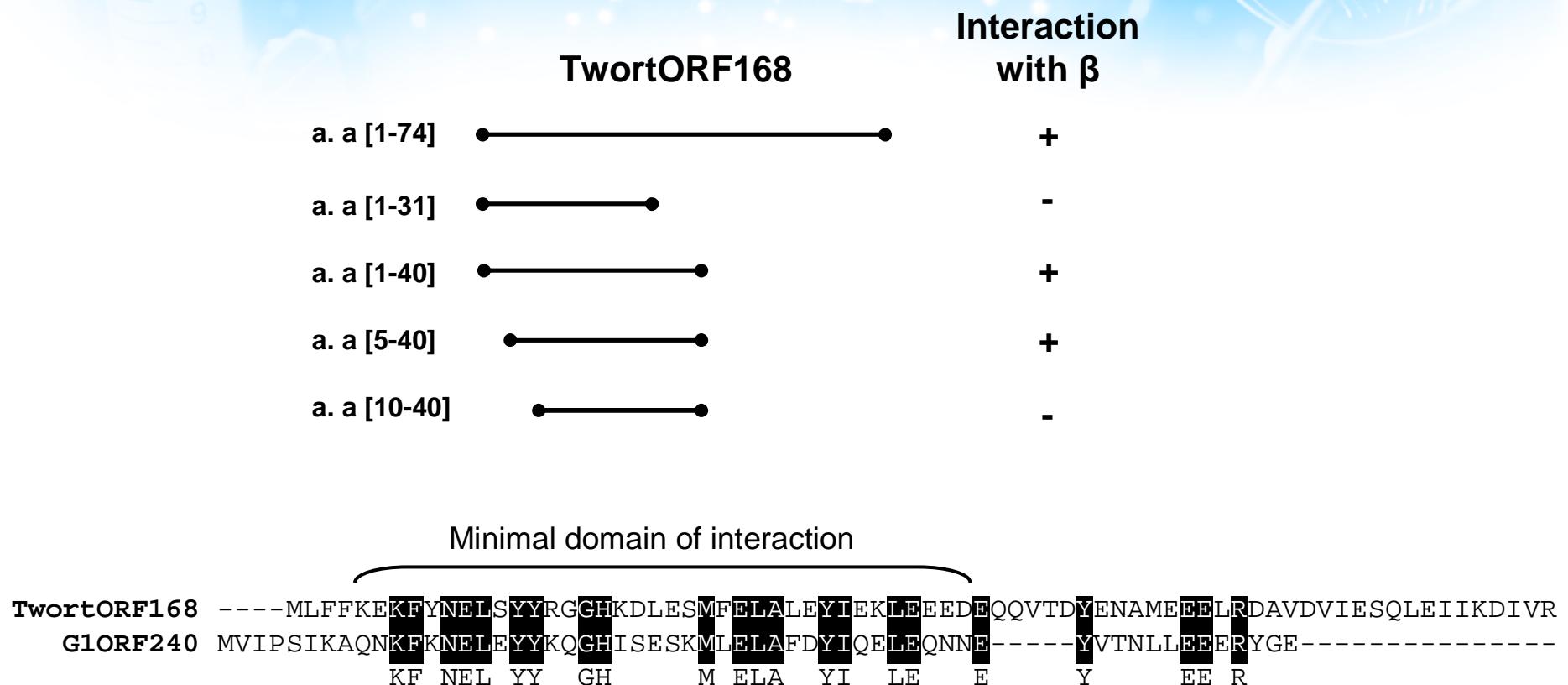
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Affinity Chromatography Identifies Replication Machinery Targets for Phage Polypeptides



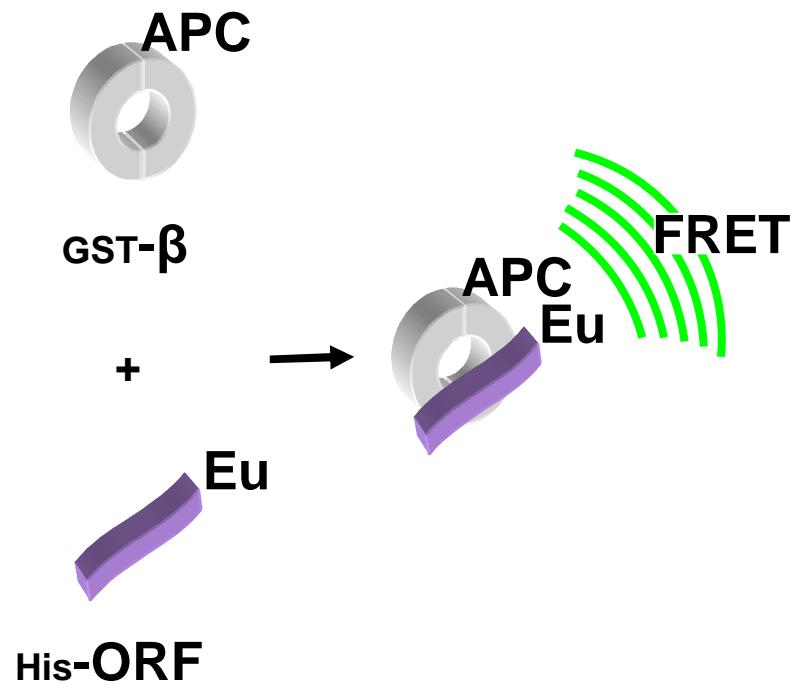
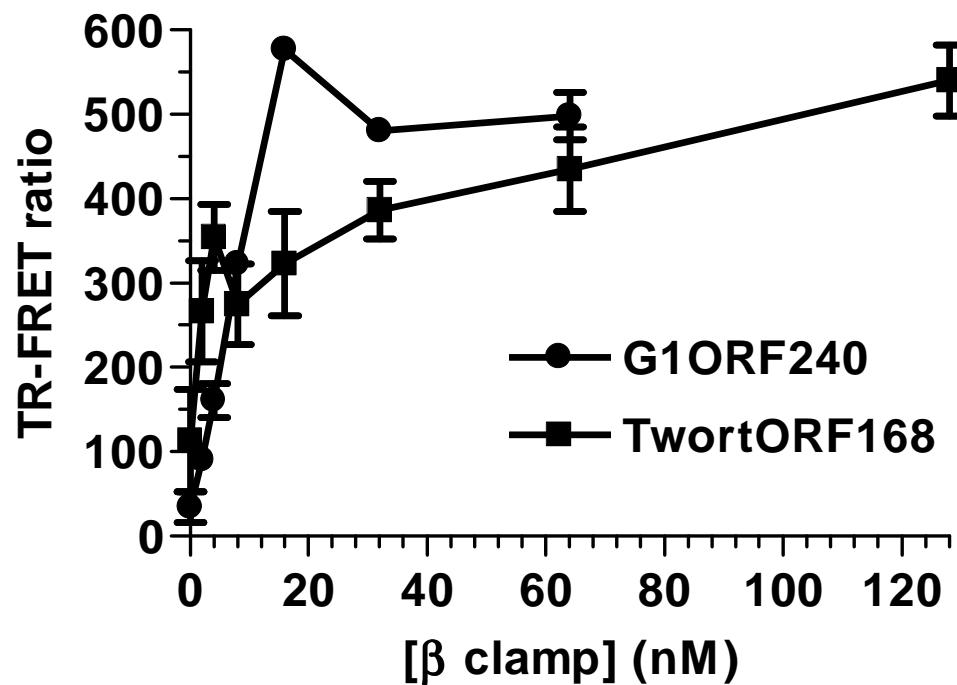
Region of Highest Sequence Similarity in ORF168 and ORF240 is Required for Binding to β

- Yeast two-hybrid assay with truncated ORF168 and full-length β → identify minimal interacting domain of phage polypeptide



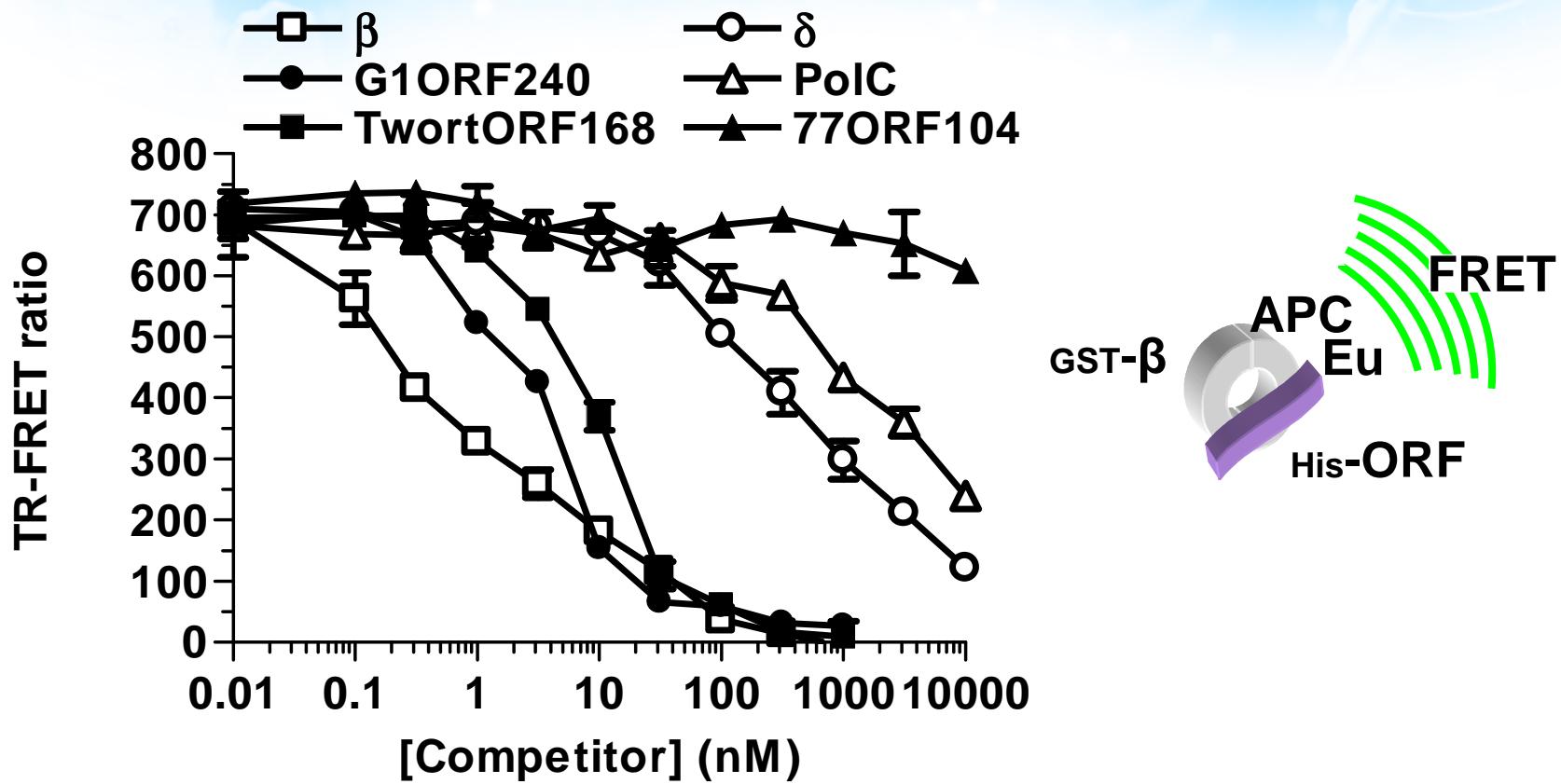
Phage Polypeptides ORF240 and ORF168 Bind to β in Solution

- TR-FRET fluorescence assay → confirm the interaction in solution:

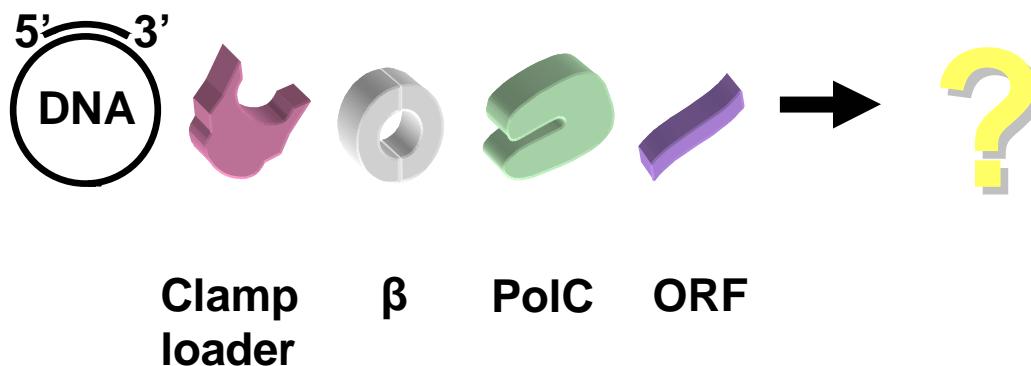
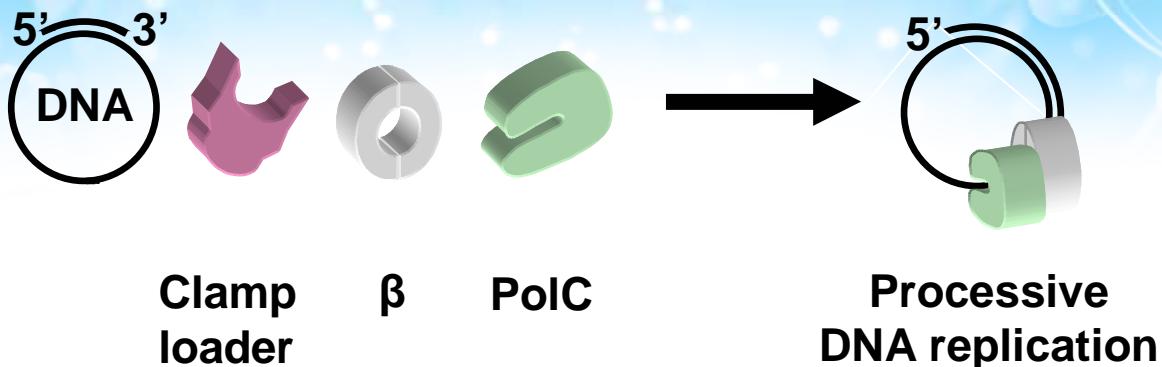


Phage Polypeptide Binding Site on β Appears to be Shared by Replicase Components

- TR-FRET fluorescence assay → study competitors of the interaction:

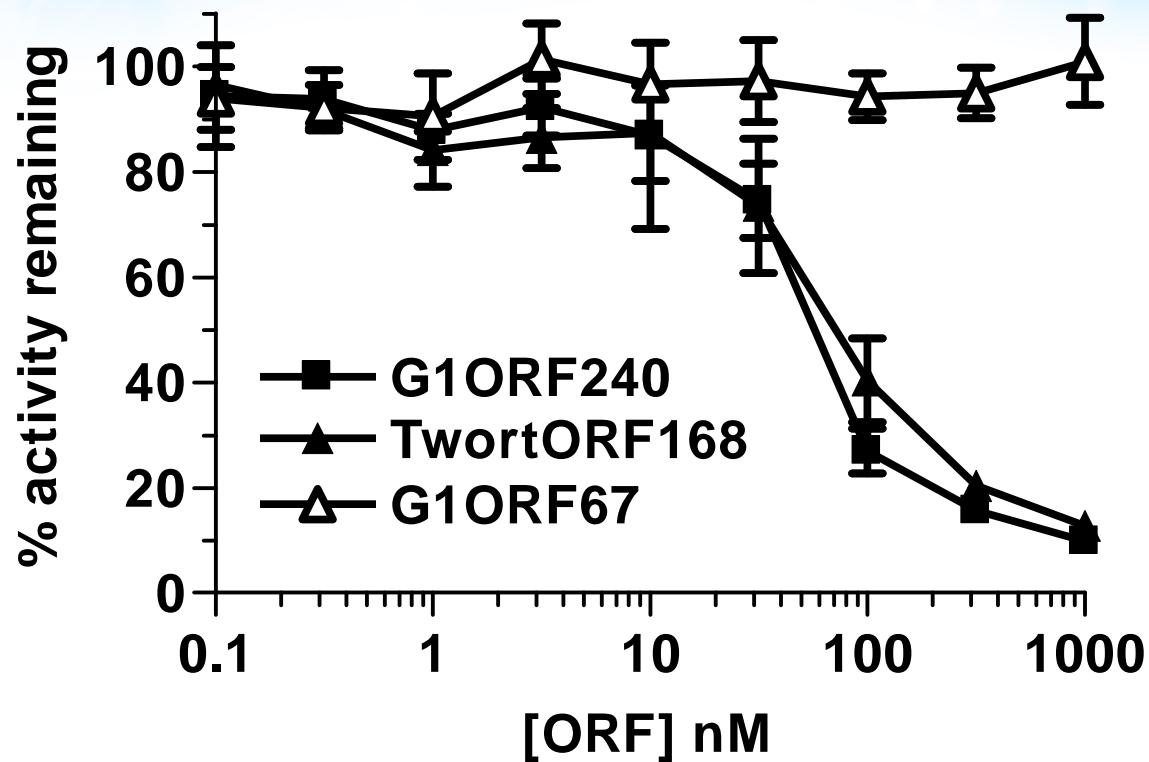


Does Phage Polypeptide Binding to β have a Functional Consequence?



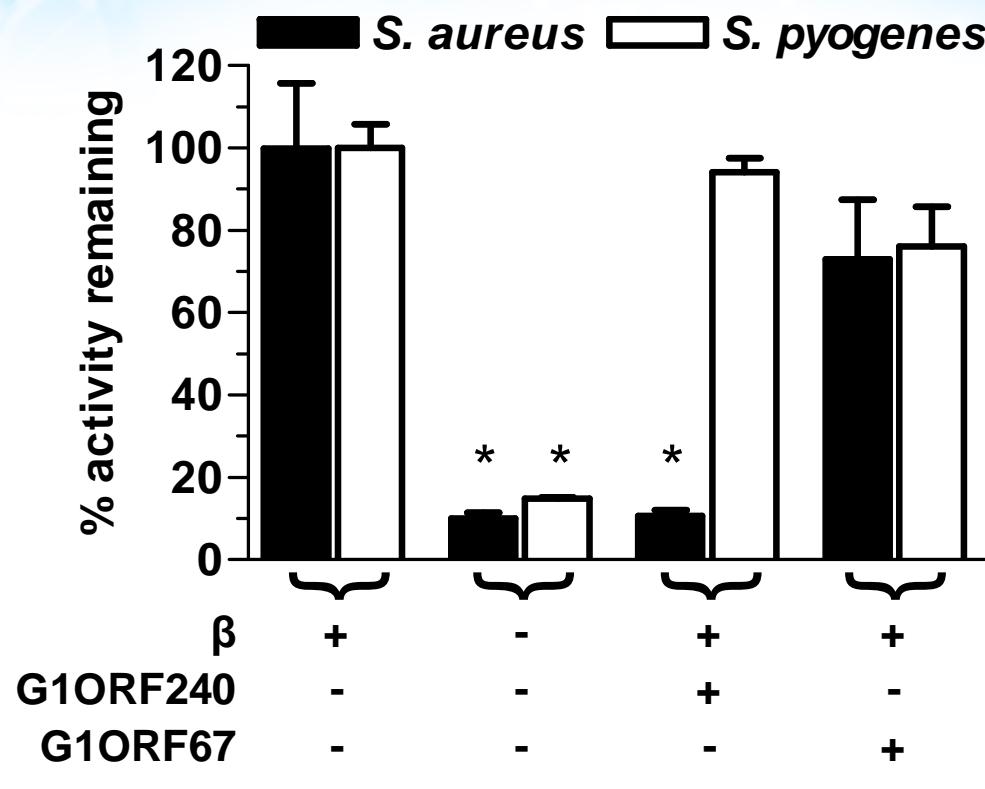
β -binding Phage Polypeptides Inhibit the *S. aureus* DNA Replicase *in vitro*

- Plate-based assay with reconstituted replicase → study effect of phage polypeptides on processive DNA synthesis *in vitro*:



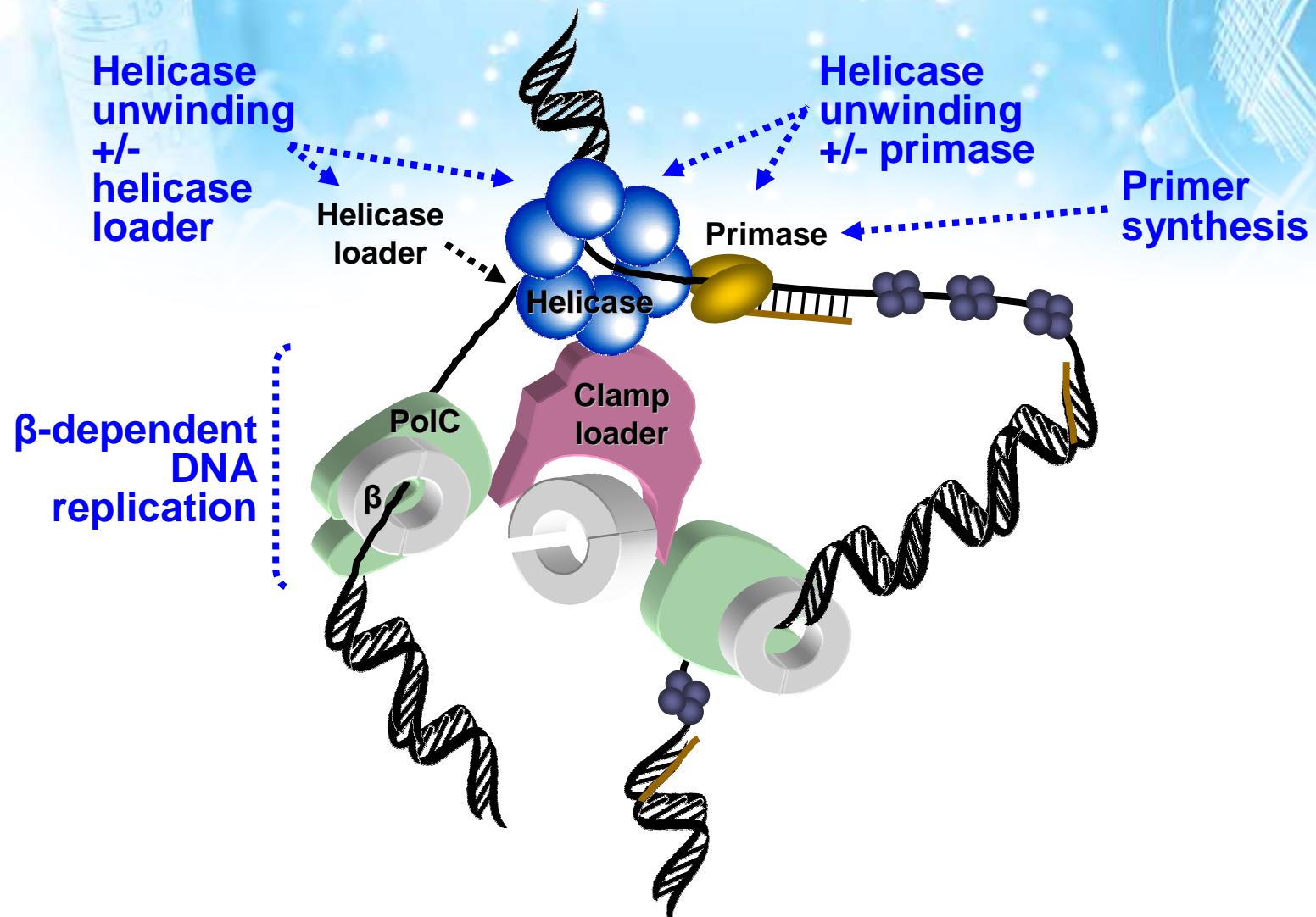
Inhibition of DNA Synthesis *In vitro* by ORF240 is Selective for *S. aureus*

- Plate-based assay with replicases from *S. aureus* and *S. pyogenes* → study selectivity of phage polypeptide inhibition of DNA synthesis *in vitro*:

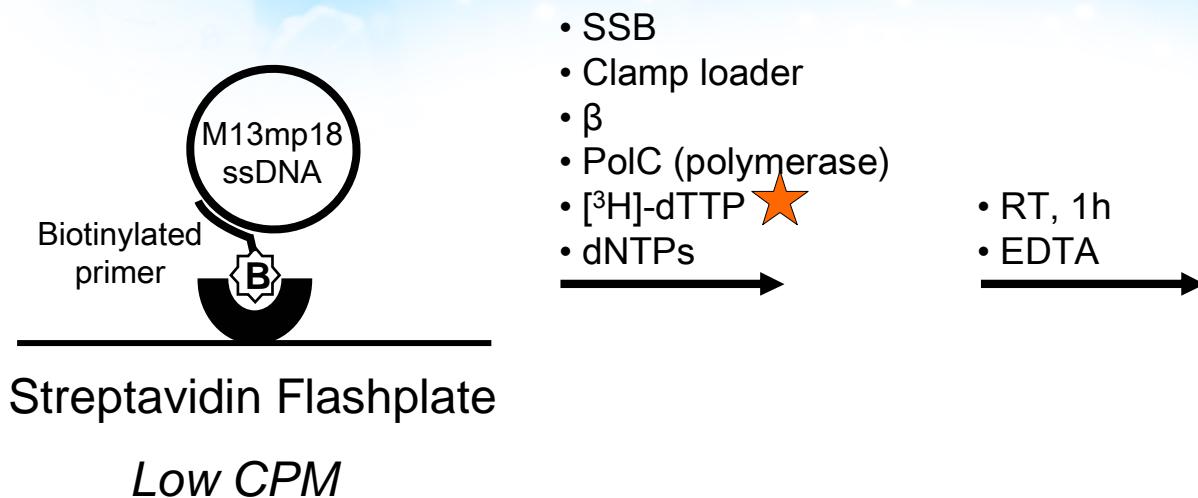


* $P < 0.05$

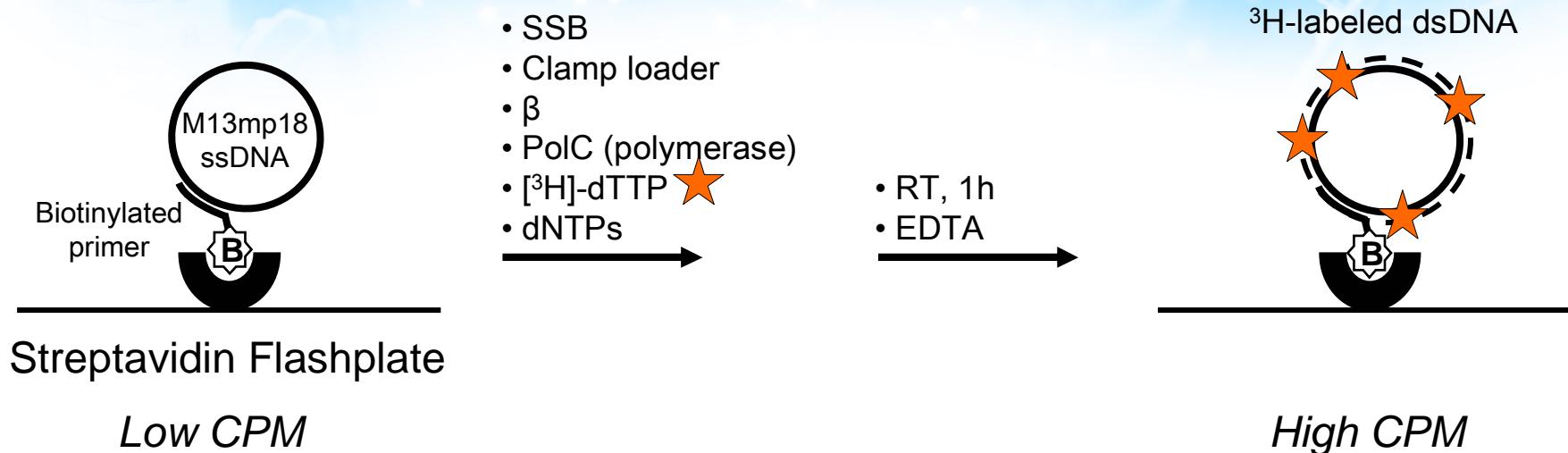
Enzyme-Based Screens for Small Molecule Inhibitors Exploit Essential Activities of the Replicase



Screening for Small-molecule Inhibitors of the *S. aureus* Replicase

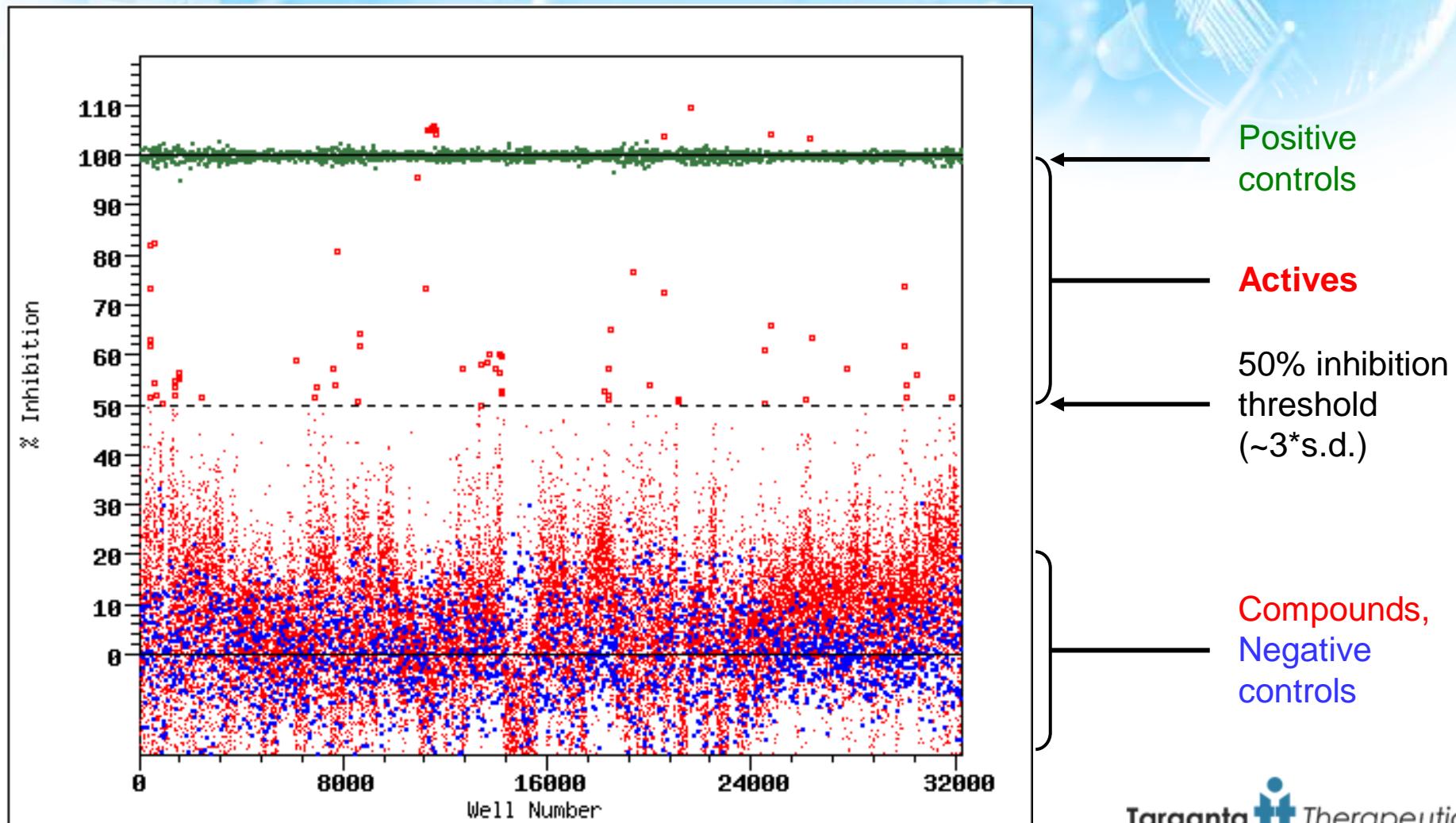


Screening for Small-molecule Inhibitors of the *S. aureus* Replicase

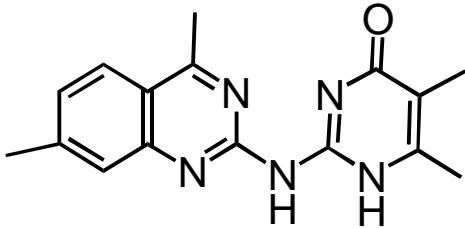
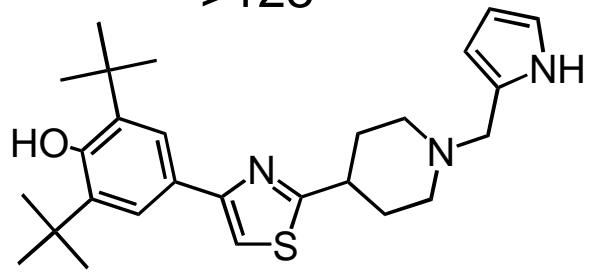


Screening for Small-molecule Inhibitors of the *S. aureus* Replicase

- HTS of 126,400 small-molecule compounds ($n=4/\text{well}$):



In vitro Activities of Two *S. aureus* Replicase Inhibitors

	Compound 1	Compound 2
IC ₅₀ (μM), <i>S. aureus</i> replicase	18 ± 7.1	7.9 ± 0.2
IC ₅₀ (μM), mammalian DNA replicase	>50	>50
IC ₅₀ (μM), DNA binding assay	>50	>50
IC ₅₀ (μM), HeLa cytotoxicity assay	>100	14
MIC (μg/mL)		
• <i>S. aureus</i> ATCC 13709 (MSSA)	4	8
• <i>S. aureus</i> ATCC 13709 + 4% HSA	64	64
• <i>M. bovis</i> BCG (Denmark, Phipps)	4	n.d.
• <i>H. influenzae</i> ATCC 49766	>32	>128
Structure		

Summary

- **ORF168 and ORF240 polypeptides:**
 - inhibit DNA synthesis selectively in *S. aureus*
 - bind selectively to the *S. aureus* DNA sliding clamp *in vitro*
 - inhibit processive DNA replication *in vitro*
- **Small-molecule inhibitors from replicase screen:**
 - active against G+ including MRSA, and efflux-deficient G-
 - validate the replicase for further inhibitor screening
 - suffer from serum shift
 - lack efficacy in rigorous models of *S. aureus* infection
- **Additional series are under study**

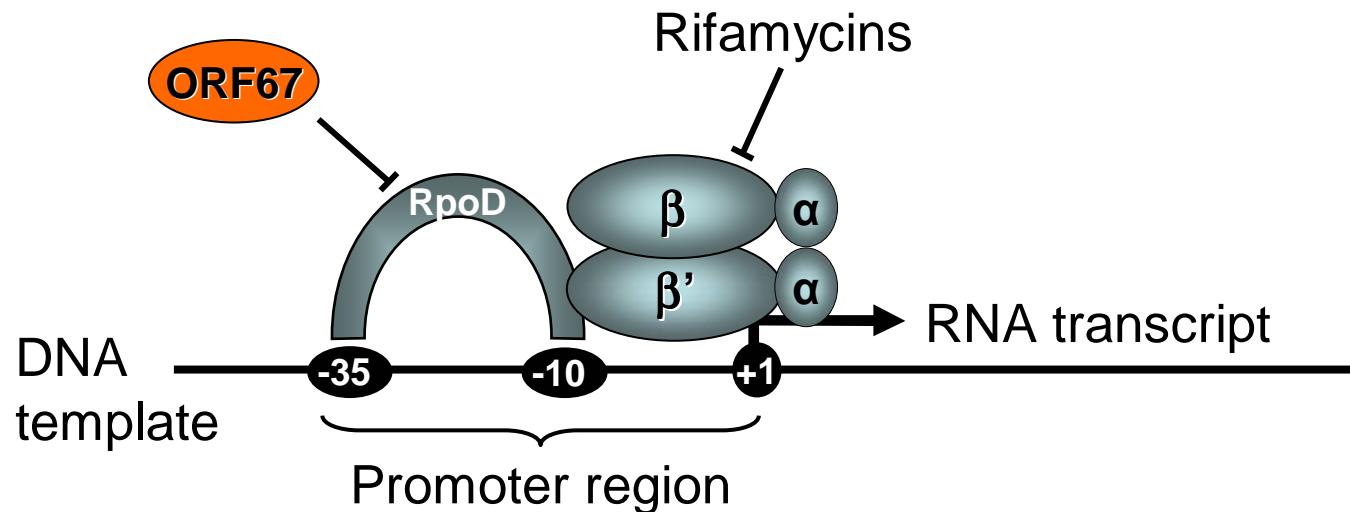


S. aureus transcription target: primary sigma factor RpoD

S. aureus RpoD is the target of phage polypeptide ORF67

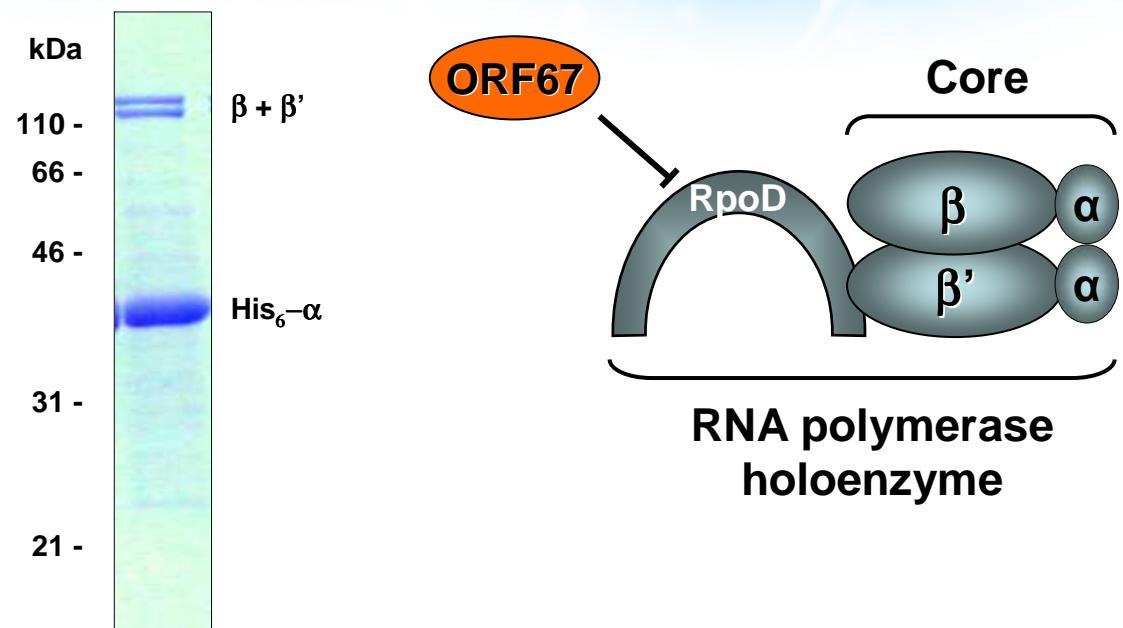
- RpoD:

- primary sigma factor in *S. aureus* transcription machinery
- ortholog of *E. coli* σ^{70}
- essential for *S. aureus* viability
- is targeted by phage polypeptide ORF67



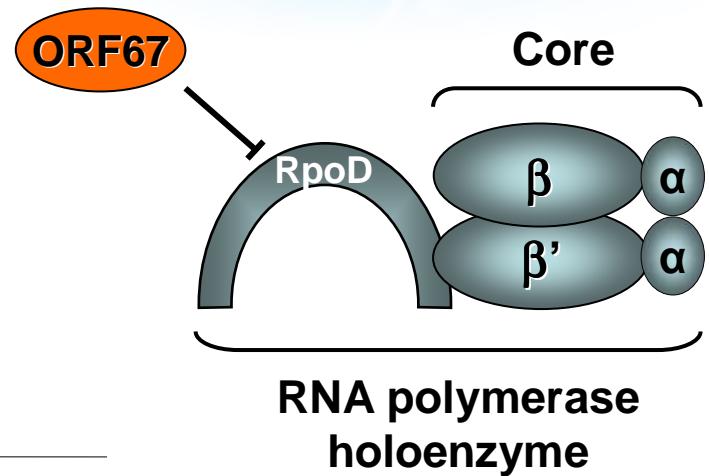
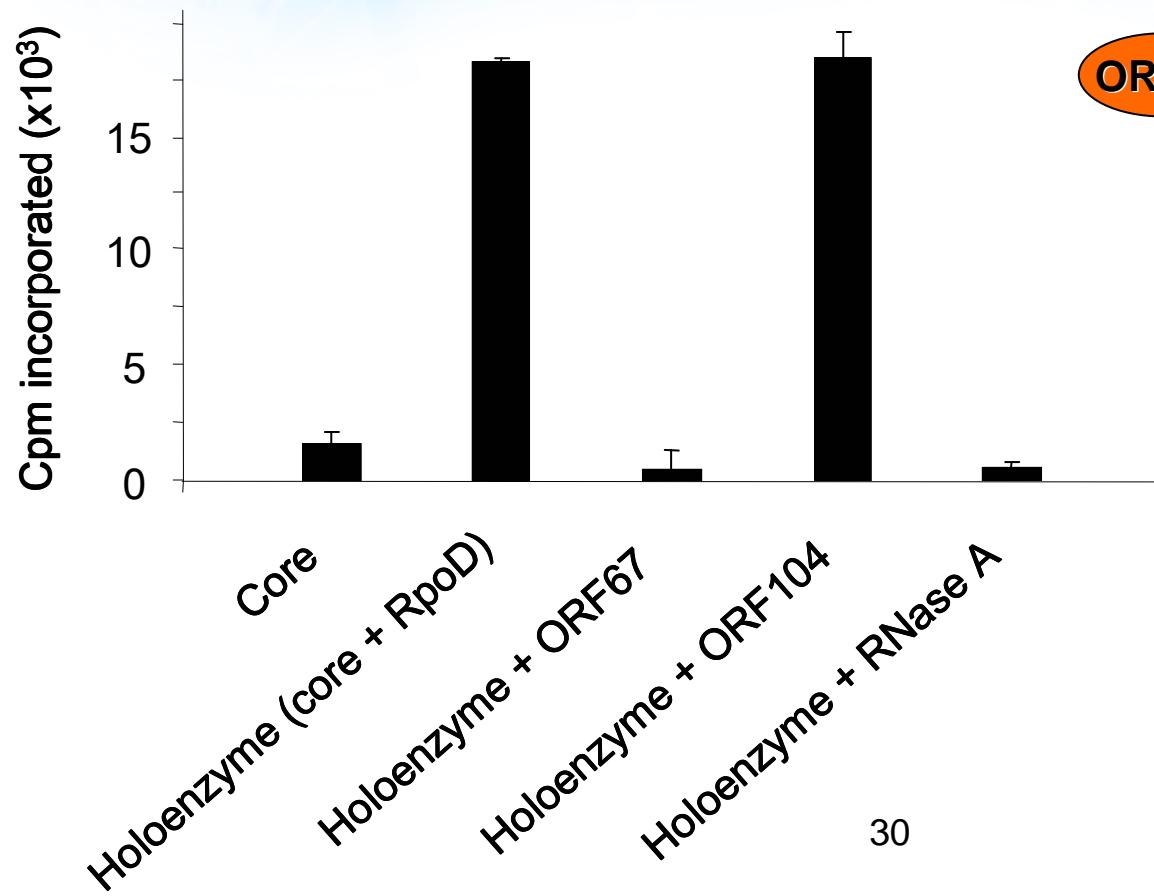
Purification of RNAP Core From *S. aureus*

- Purification of RNAP from *S. aureus* using His-tagged α subunit → develop *in vitro* assay for RNA synthesis:

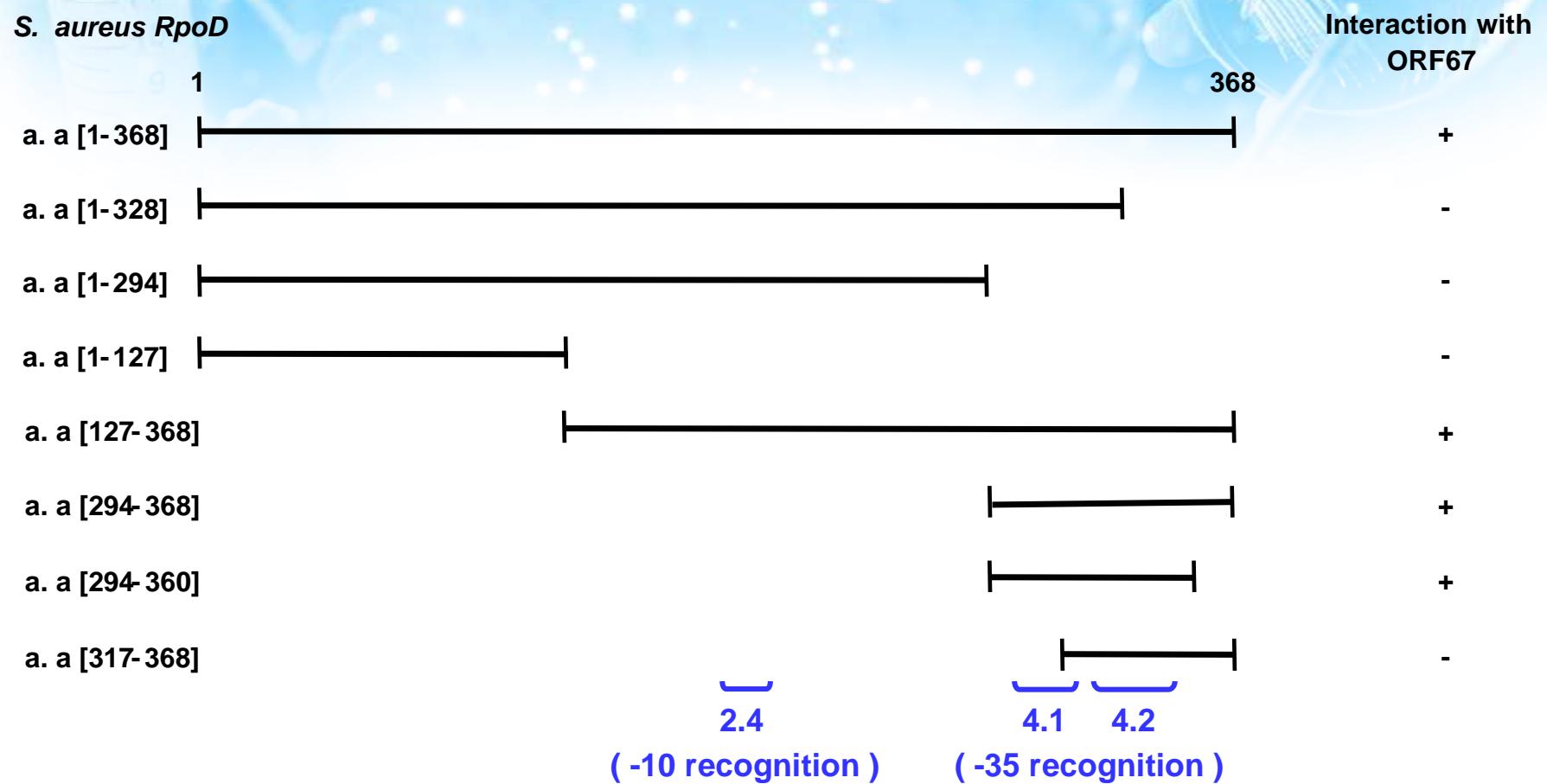


RpoD-Specific Phage Polypeptide (ORF67) Inhibits *S. aureus* *in vitro* Transcription

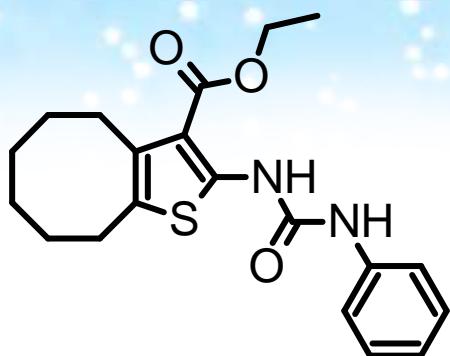
- Plate-based assay with purified *S. aureus* RNAP → study effect of phage polypeptides on RNA synthesis *in vitro* and conduct HTS:



Regions 4.1 and 4.2 of *S. aureus* RpoD Comprise the Minimal Domain For ORF67 Interaction

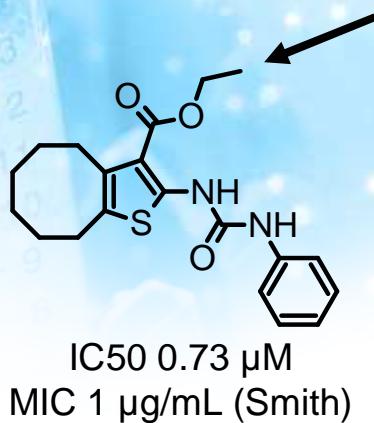


Transcription Screen Identifies a Novel Ureidothiophene Carboxylate Inhibitor

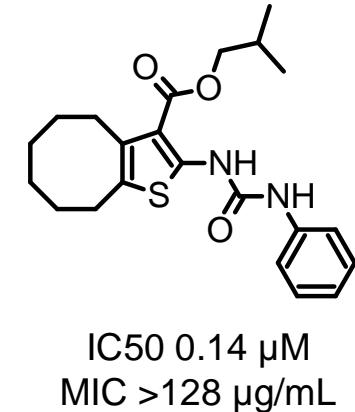
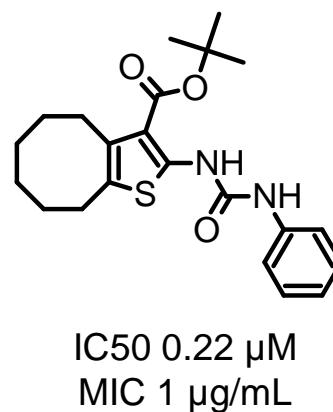
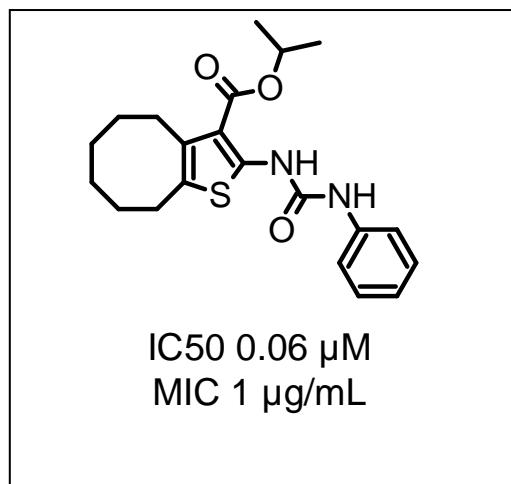
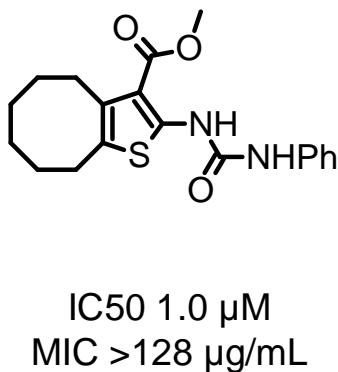


⇒ IC50 (<i>in vitro</i> <i>S. aureus</i> transcription):	0.73 μM
⇒ MIC (<i>S. aureus</i> Smith ATCC 13709):	1 μg/mL
⇒ MIC (50% serum):	>128 μg/mL
⇒ Spectrum:	limited to Staphylococci
⇒ IC50 (<i>in vitro</i> HeLa cytotoxicity):	>100 μM
⇒ IC50 (<i>E. coli</i> <i>in vitro</i> transcription):	>100 μM
⇒ IC50 (mammalian <i>in vitro</i> transcription):	>100 μM

Ester Variations and Activity

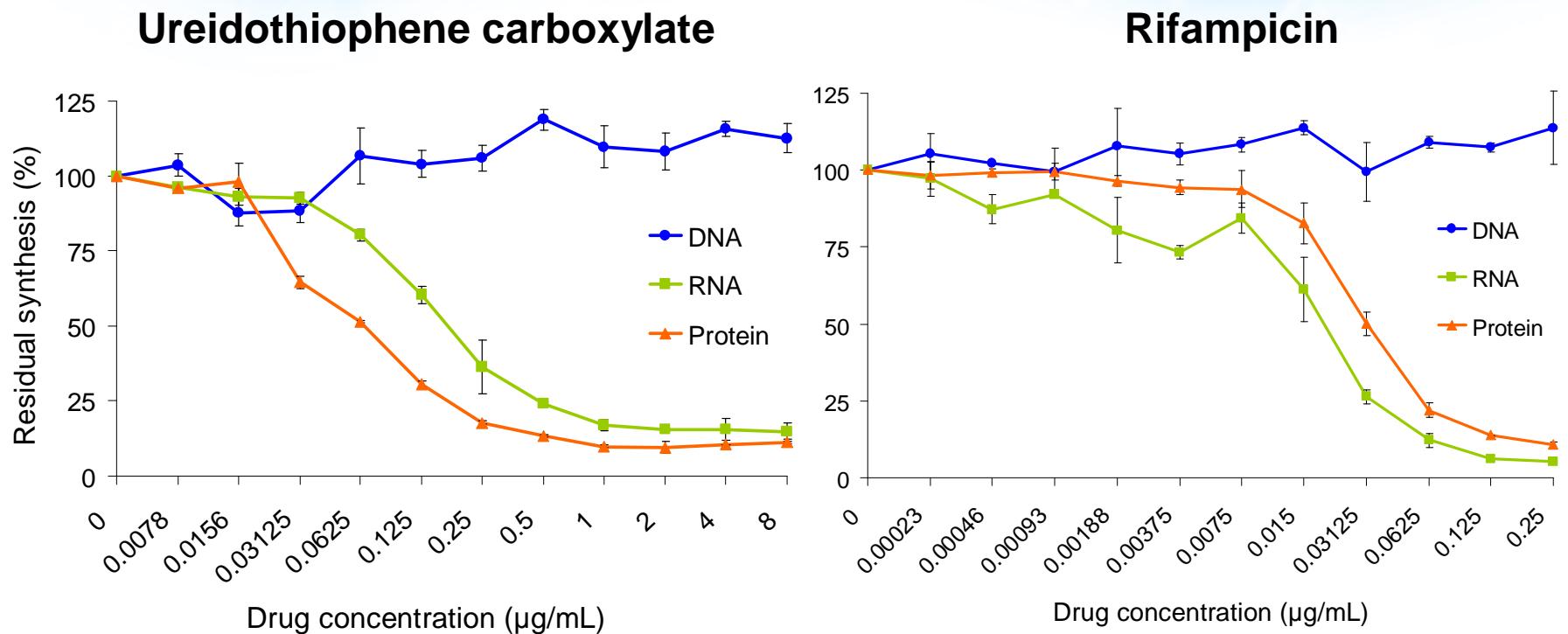


- Ester functionality is necessary
 - amides, ketones, alcohols, acid tested
- Polar groups are undesirable
 - small heterocycles +/- charge tested
- **Isopropyl ester is optimum:**



Ureidothiophene Carboxylate Inhibits Transcription in Growing *S. aureus* Cells

- Macromolecular synthesis assay in *S. aureus* → Ureidothiophene carboxylate inhibits RNA and protein synthesis similarly to Rifampicin:

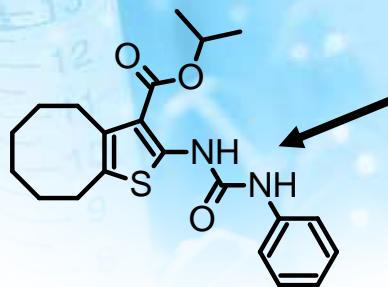


Ureidothiophene Carboxylate is Active Against Antibiotic Resistant Strains of *S. aureus*

Resistant Category	n	MIC or MIC range ($\mu\text{g/mL}$)
Mupirocin-resistant	12	0.5 (11 strains) >128 (1 strain)
Rifampicin-resistant	9	< 0.125 - 1
MRSA	14	0.25 - 2
VISA ATCC 700699	1	0.25

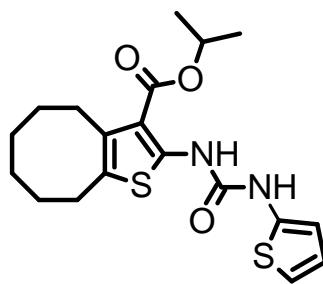
- Activity against Rif^R strains suggests distinct binding site or mechanism

Urea Variations and Activity

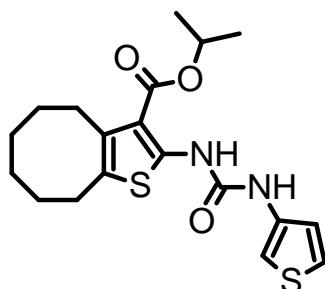


IC₅₀ 0.06 μM
MIC 0.5 μg/mL (Smith)

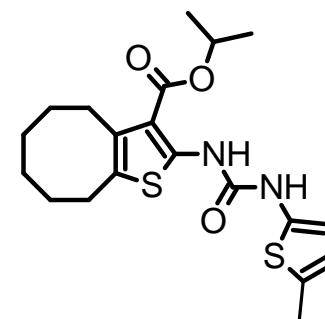
- Urea functionality is necessary; amides, carbamates, thioureas, sulfuric diamides lose inhibitory activity
- Replacement of phenyl ring with alicyclics or heterocyclics abolishes antibacterial activity
- Substituents on phenyl group abolish antibacterial activity
 - meta, para substituents retain inhibitory activity
 - ortho substituents destroy inhibitory activity
- **Only thiophenes are tolerated as phenyl replacements:**



IC₅₀ 0.06 μM
MIC 0.5 μg/mL

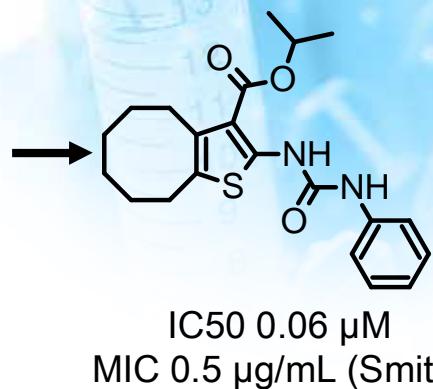


IC₅₀ 0.20 μM
MIC 0.5 μg/mL

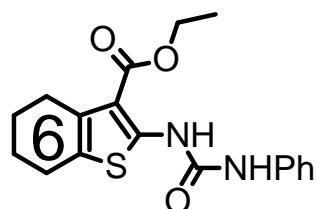


IC₅₀ 0.49 μM
MIC 1 μg/mL

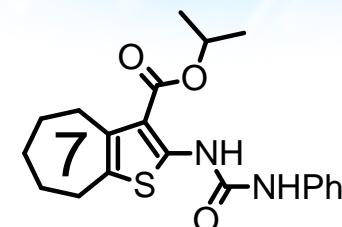
Ring Variations and Activity



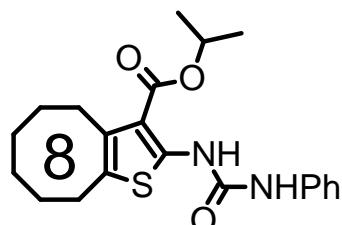
- Heteroatoms in ring abolish antibacterial activity
- Acyclic replacements are detrimental (IC₅₀ 5-10 μM)
- **Eight and nine membered rings optimum:**



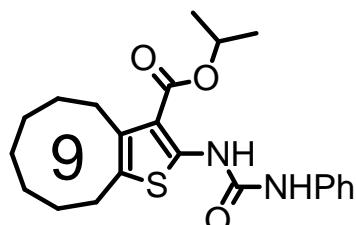
IC₅₀: 2.4 μM
MIC >128 μg/mL



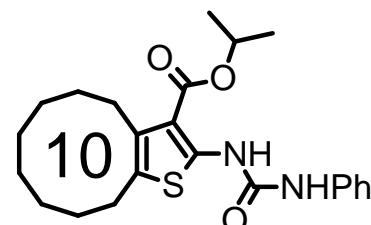
IC₅₀: 0.1 μM
MIC 1 μg/ml



IC₅₀: 0.06 μM
MIC 0.5 μg/mL

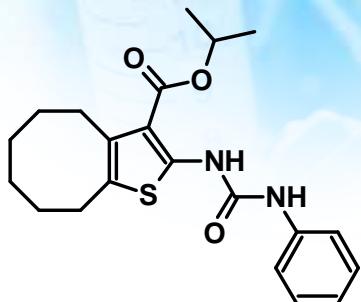


IC₅₀: 0.05 μM
MIC 0.5 μg/mL



IC₅₀: 0.14 μM
MIC >128 μg/mL

Ureidothiophene Carboxylate - Summary



IC₅₀ 0.06 μM
MIC 0.5 μg/mL

- >120 compounds made in 6 month campaign
- Compound is likely only active against staphylococci and is subject to high frequencies of resistance
- Well-tolerated in mice at 2x25 mg/kg i.v. bolus
- Active in a low-stringency mouse model of systemic *S. aureus* infection (i.p. infection / i.p. injection)
- Additional SAR of hydrophobic ring required to address serum binding issue

Acknowledgements

Targanta's CSO

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Targanta Biology

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